

# Isoprostanes

L. Jackson Roberts, II<sup>1</sup> and Ginger L. Milne

Division of Clinical Pharmacology, Vanderbilt University Medical Center, Nashville, TN

**Abstract** The isoprostanes (IsoPs) are a unique series of prostaglandin-like compounds formed *in vivo* via a nonenzymatic mechanism involving the free radical-initiated peroxidation of arachidonic acid. This article summarizes our current knowledge of these compounds. Herein, a historical account of their discovery and the mechanism of their formation are described. A specific class of IsoPs, the F<sub>2</sub>-IsoPs, are stable, robust molecules that can be measured as indices of endogenous oxidant stress. The utility of these molecules as biomarkers and methods by which these compounds can be quantified are discussed. In addition to the F<sub>2</sub>-IsoPs, isoprostanes with other prostane ring structures as well as oxidation products with furan and dioxolane rings can be generated from arachidonic acid. And, in more recent years, isoprostane-like compounds have been shown to be formed from polyunsaturated fatty acids including eicosapentaenoic acid [C20:5, ω-3], docosahexaenoic acid [C22:6, ω-3], and adrenic acid [C22:4, ω-6]. These findings will be summarized as well.—Roberts, L. J., II, and G. L. Milne. *Isoprostanes*. *J. Lipid Res.* 2009. 50: S219–S223.

**Supplementary key words** oxidative stress • lipid peroxidation • prostaglandins

Free radicals derived primarily from molecular oxygen have been implicated in a variety of human conditions and disorders, including atherosclerosis and associated risk factors, which include smoking and obesity, ischemia/reperfusion injury, and neurodegenerative diseases such as Alzheimer's disease and Huntington's disease (1). Lipids are readily attacked by free radicals resulting in the formation of a number of peroxidation products. One class of oxidation products formed in abundance *in vitro* and *in vivo* is the isoprostanes (IsoPs), which were discovered by our laboratory in 1990. IsoPs are a series of prostaglandin (PG)-like compounds produced by the free radical-catalyzed peroxidation of arachidonic acid independent of the cyclooxygenase (2). Over the past 20 years, we and others (3) have carried out a large number of studies defining the basic chemistry and biochemistry involved in the

formation and metabolism of the IsoPs. In addition, we have shown that levels of IsoPs are increased in a number of human diseases, and it is currently recognized that measurement of these molecules is the most accurate analytical method to assess oxidative injury *in vivo*. Further, a number of IsoPs have been found to possess potent biological activity and thus are likely also mediators of oxidant injury (4). In recent years, additional related compounds, derived from various polyunsaturated fatty acids such as eicosapentaenoic acid (EPA) (5), docosahexaenoic acid (DHA) (6), and, more recently, adrenic acid (7), have been discovered to be formed as products of the IsoP pathway. It is the purpose herein to summarize our current knowledge regarding the IsoPs including the chemistry and biochemistry of their formation, the utility of measuring these compounds as markers of *in vivo* oxidant stress, and their pharmacological properties.

## DISCOVERY OF F<sub>2</sub>-ISOPROSTANES

In 1990, Morrow et al. (2) discovered the first class of IsoPs, the F<sub>2</sub>-IsoPs, so named because they contain F-type prostane rings analogous to PGF<sub>2α</sub>. A mechanism to explain the formation of the F<sub>2</sub>-IsoPs from arachidonic acid is outlined in **Fig. 1**. Based on this mechanism of formation, four F<sub>2</sub>-IsoP regioisomers are generated; compounds are denoted as 5-, 12-, 8-, or 15-series regioisomers, depending on the carbon atom to which the side chain hydroxyl is attached (8). An alternative nomenclature system for the IsoPs has been proposed by Rokach et al. (9) in which the abbreviation iP is used for isoprostane, and the regioisomers are denoted as III–VI based upon the number of carbons between the omega carbon and the first double bond.

An important structural distinction between IsoPs and cyclooxygenase (COX)-derived PGs is that the former contain side chains that are predominantly oriented *cis* to the prostane ring while the latter possess exclusively *trans* side

Abbreviations: COX, cyclooxygenase; DHA, docosahexaenoic acid; EPA, eicosapentaenoic acid; GC, gas chromatography; GSH, glutathione; IsoF, isofuran; IsoK, isoketal; IsoP, isoprostane; LC, liquid chromatography; MS, mass spectrometry; NICI, negative ion chemical ionization; NP, neuroprostane; PG, prostaglandin.

<sup>1</sup>To whom correspondence should be addressed.

e-mail: jack.roberts@vanderbilt.edu

Supported by National Institutes of Health Grants GM15431, GM42056, AG023597, DK48831, ES13125 and ES00267.

Manuscript received 9 October 2008.

Published, *JLR Papers in Press*, October 28, 2008.  
DOI 10.1194/jlr.R800037-JLR200

Copyright © 2009 by the American Society for Biochemistry and Molecular Biology, Inc.

This article is available online at <http://www.jlr.org>

## QUANTIFICATION OF F<sub>2</sub>-ISOPs AS AN INDEX OF ENDOGENOUS OXIDANT STRESS

A true utility of the F<sub>2</sub>-IsoPs is in the quantification of lipid peroxidation and thus oxidant stress status in vivo (19, 20). Because they are stable molecules, measurement of the F<sub>2</sub>-IsoPs has revolutionized the ability to quantify oxidative injury in vivo. In a recent multi-investigator study, termed the Biomarkers of Oxidative Stress Study, sponsored by the National Institute of Health, it was found that the most accurate method to assess in vivo oxidant stress status is the quantification of plasma or urinary IsoPs, and thus, currently, quantification of these compounds provides the “gold standard” to assess oxidative injury in vivo (3).

A number of methods have been developed to quantify the IsoPs. Our laboratory uses a gas chromatographic/negative ion chemical ionization mass spectrometric (GC/NICI-MS) approach employing stable isotope dilution (21). For quantification purposes, we measure the F<sub>2</sub>-IsoP, 15-F<sub>2t</sub>-IsoP, and other F<sub>2</sub>-IsoPs that coelute with this compound. Several internal standards are available from commercial sources to quantify the IsoPs. The advantages of MS over other approaches include its high sensitivity and specificity, which yield quantitative results in the low picogram range. Its drawbacks are that it is labor intensive and requires considerable expenditures on equipment.

Several alternative GC/MS assays have been developed by different investigators including Pratico et al. (22) that quantify other IsoP regioisomers and Mas et al. (23) who utilize an F-ring IsoP deriving from DHA, termed 4-F<sub>4t</sub>-NP, as an internal standard. In addition, a number of liquid chromatographic (LC)/MS methods for F<sub>2</sub>-IsoPs have been developed (24–26). One advantage of LC/MS methods is that the sample preparation for analysis is simpler than that for GC/MS because no derivatization of the molecule is required.

Alternative methods have also been developed to quantify IsoPs using immunological approaches (27). Antibodies have been generated against 15-F<sub>2t</sub>-IsoP and several immunoassay kits are commercially available. A potential drawback of these methods is that limited information is currently available regarding their precision and accuracy. In addition, little data exist comparing IsoP levels determined by immunoassay to MS. Despite potential limitations, immunoassays have expanded IsoP research due to their low cost and relative ease of use (20).

Normal levels of F<sub>2</sub>-IsoPs in healthy humans have been defined (4, 28). Defining these levels is particularly important in that it allows for an assessment of the effects of diseases on endogenous oxidant tone and allows for the determination of the extent to which various therapeutic interventions affect levels of oxidant stress. Elevations of IsoPs in human body fluids and tissues have been found in a diverse array of human disorders, including atherosclerosis (29), diabetes (30), obesity (31), cigarette smoking (32), neurodegenerative diseases (33), and many others. Further, treatments for some of these conditions, including antioxidant supplementation, antidiabetic treatments, cessation of smoking, and even weight loss, have been shown to decrease production of F<sub>2</sub>-IsoPs (34, 35).

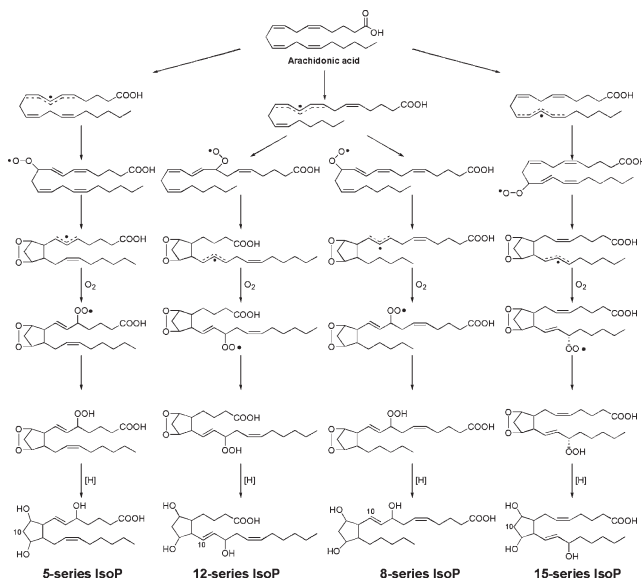


Fig. 1. Pathway of formation of F<sub>2</sub>-isoprostanes. IsoP, isoprostane.

chains (2). In this regard, however, we have reported that PGs can be formed via the IsoP pathway as smaller amounts of endoperoxides containing *trans* side chains are generated in vitro and in vivo by this mechanism (10). In this case, PGs derived via the IsoP pathway can be distinguished from those formed by COX because the former are generated as a racemic mixture while the latter are enantiomerically pure. A second important difference between IsoPs and PGs is that IsoPs are formed primarily in situ esterified to phospholipids and are subsequently released by a phospholipase(s) (11), while PGs are generated only from free arachidonic acid. Molecular modeling of IsoP-containing phospholipids reveals them to be remarkably distorted molecules (12). Thus, the formation of these abnormal phospholipids would be expected to exert profound effects on membrane fluidity and integrity, well-known sequelae of oxidant injury.

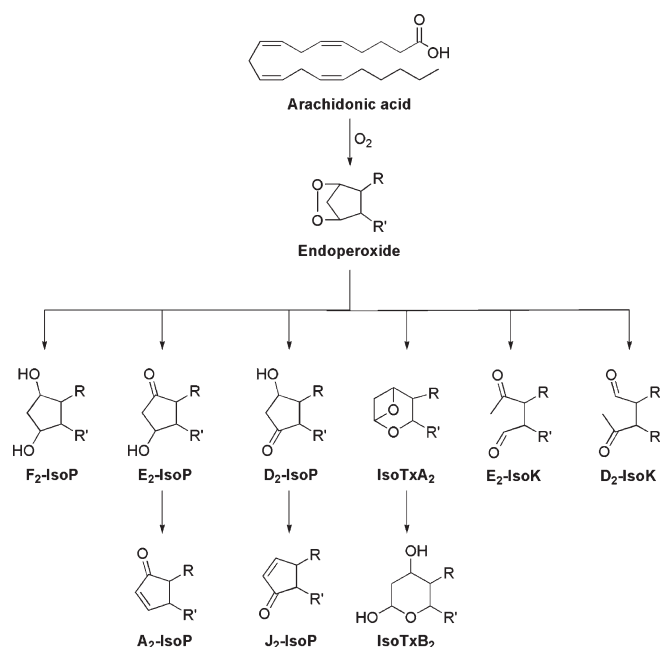
In this regard, the biological activity of the F<sub>2</sub>-IsoPs has been studied. While these molecules are formed in situ on phospholipids as discussed, studies exploring the bioactivity of IsoPs have been performed using unesterified compounds as free acids. One F<sub>2</sub>-IsoP that is produced abundantly in vivo and has been extensively tested for biological activity is 15-F<sub>2t</sub>-IsoP (8-iso-PGF<sub>2α</sub>) (13). This IsoP has been found to be a potent vasoconstrictor in a variety of vascular beds, including the kidney, lung, heart, brain, and others (14, 15). In addition, 15-F<sub>2t</sub>-IsoP induces endothelin release and proliferation of vascular smooth muscle cells. There is also additional evidence that this molecule can increase resistance to aspirin inhibition of platelet aggregation in platelets as well as inhibit platelet aggregation in human whole blood (16). These vasoactive and platelet effects of 15-F<sub>2t</sub>-IsoP have been shown to result from interaction with the thromboxane receptor, a G-protein coupled transmembrane eicosanoid receptor, based on the finding that these effects could be abrogated by thromboxane receptor antagonists (17, 18).

## FORMATION OF ISOPs WITH ALTERNATIVE RING STRUCTURES

Since the initial discovery of the F<sub>2</sub>-IsoPs, the laboratories of Roberts and Morrow have shown that the IsoP pathway provides a mechanism for the generation of other classes of IsoPs from arachidonic acid, which differ in regards to the functional groups on the prostane ring. The classes of all known IsoPs are shown in **Fig. 2**. In addition to undergoing reduction to yield F<sub>2</sub>-IsoPs, the arachidonoyl endoperoxide intermediate can undergo isomerization to yield E- and D-ring IsoPs, which are isomeric to PGE<sub>2</sub> and PGD<sub>2</sub>, respectively (36). E<sub>2</sub>/D<sub>2</sub>-IsoPs are formed competitively with F<sub>2</sub>-IsoPs, and studies have demonstrated that the depletion of cellular reducing agents, such as glutathione (GSH) or α-tocopherol, favors the formation of E<sub>2</sub>/D<sub>2</sub>-IsoPs over that of reduced F<sub>2</sub>-IsoPs (37).

E<sub>2</sub>/D<sub>2</sub>-IsoPs are not terminal products of the IsoP pathway. These compounds readily dehydrate in vivo to yield A<sub>2</sub>/J<sub>2</sub>-IsoPs, which are also known as cyclopentenone IsoPs because they contain an α,β-unsaturated cyclopentenone ring structure (38). A<sub>2</sub>/J<sub>2</sub>-IsoPs are highly reactive electrophiles, which readily form Michael adducts with cellular thiols, including those found on cysteine residues in proteins and GSH (39). These cyclopentenone IsoPs are rapidly metabolized in vivo by glutathione transferase enzymes to water-soluble modified glutathione conjugates (40). A major urinary cyclopentenone IsoP metabolite in rats, a 15-A<sub>2</sub>-IsoP mercapturic acid sulfoxide conjugate, has been identified (40).

Thromboxane-like molecules have also been reported to be generated via the IsoP pathway, although compounds with structures similar to prostacyclin have not been reported. Further, a series of compounds termed isoketals

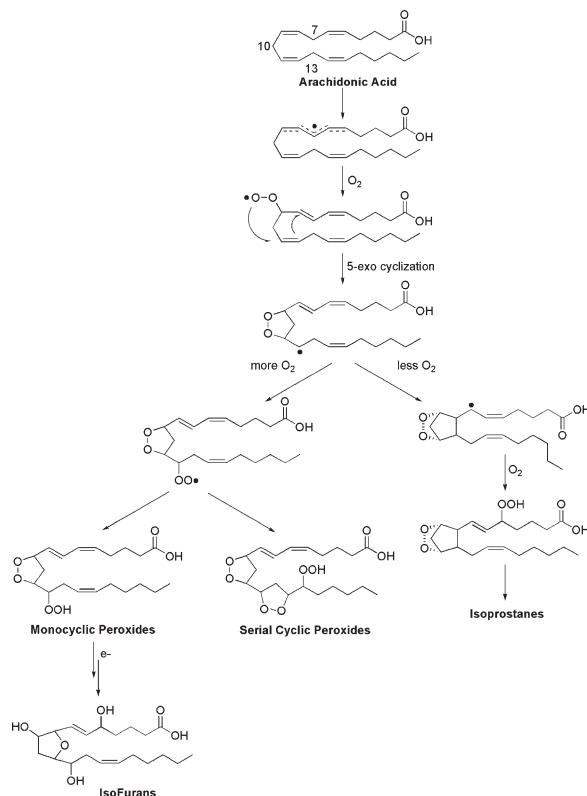


**Fig. 2.** Oxidation of arachidonic acid to yield isoprostanes of differing ring structures. IsoK, isoketal.

(IsoKs or isolevuglandins) can be generated via the IsoP pathway and result from opening of the cyclopentane ring (41). These compounds are highly reactive and readily adduct lysine residues on proteins resulting in covalent modification and protein dysfunction and cross-linking.

## FORMATION OF OTHER PEROXIDATION PRODUCTS FROM ARACHIDONIC ACID

In addition to being able to form PG-like molecules, arachidonic acid can be oxidized to yield molecules with cyclic peroxide and furan ring structures. These molecules are formed in competition with isoprostanes from the key intermediate 1 as can be seen in **Fig. 3**. This radical intermediate can either undergo a 5-exo cyclization reaction, which does not require oxygen, to yield IsoPs or react with molecular oxygen to yield monocyclic peroxides and serial cyclic peroxides (42), which contain cyclic peroxide ring structures, or compounds termed isofurans (IsoFs), which contain a substituted tetrahydrofuran ring (43). Based upon this mechanism of formation, one would hypothesize that as oxygen tension increases, the formation of cyclic peroxides and IsoFs would be favored compared with IsoPs. Indeed our laboratory has shown that when arachidonic acid is oxidized at differing oxygen tensions (1% O<sub>2</sub>, 21% O<sub>2</sub>, and 100% O<sub>2</sub>) levels of IsoFs increase with oxygen tension, while no significant increase in IsoPs is observed between 21% O<sub>2</sub> and 100% O<sub>2</sub> (44).



**Fig. 3.** Oxidation of arachidonic acid to yield isoprostanes, mono and serial cyclic peroxides, and isofurans.

Considering these findings, IsoFs may provide a better index of oxidant stress than IsoPs in conditions associated with higher oxygen levels. Indeed, in an experimental setting of hyperoxia-induced lung injury in mice, we found that IsoFs esterified in the lung tissues of these animals are significantly increased compared with controls while F<sub>2</sub>-IsoPs remained unchanged (44). Further, in a study in which rats were administered kainic acid, it was found that seizures induced increases F<sub>2</sub>-IsoPs and IsoFs in a manner dependent upon cellular levels of O<sub>2</sub> in the hippocampus (45). F<sub>2</sub>-IsoPs were increased in the peak hypoxia phase of the seizure while IsoFs were increased in the "reoxygenation" phase of the seizure.

#### FORMATION OF ISOPs FROM OTHER POLYUNSATURATED FATTY ACIDS

Arachidonic acid is not the only polyunsaturated fatty acid that can be oxidized to generate IsoPs. F-ring IsoPs have been shown to be generated from the peroxidation of the omega-3 polyunsaturated fatty acids EPA [C20:5, ω-3, F<sub>3</sub>-IsoPs] and DHA [C22:6, ω-3, F<sub>4</sub>-neuroprostanes (NPs)] (5, 6). Our interest in examining the formation of IsoP-like compounds from EPA and DHA stems from emerging evidence that has implicated increased dietary intake of fish oil, which contains large amounts of EPA and DHA, as being beneficial in the prevention and treatment of a number of diseases, including atherosclerotic cardiovascular disease and sudden death, neurodegeneration, and various inflammatory disorders, among others. Further, recent data have suggested that the anti-inflammatory effects and other biologically relevant properties of ω-3 fatty acids are due, in part, to the generation of various bioactive oxidation products (46). We thus hypothesized that EPA- and DHA-derived IsoPs could contribute to the beneficial biological effects of fish oil supplementation. Indeed, one report states that the EPA-derived IsoP, 15-F<sub>3t</sub>-IsoP, possesses activity that is different from 15-F<sub>2t</sub>-IsoP in that it does not affect human platelet shape change or aggregation (47).

Interestingly, our laboratory has found that the levels of IsoPs generated from the oxidation of EPA significantly exceeds those of F<sub>2</sub>-IsoPs generated from arachidonic acid, perhaps because EPA contains more double bonds and is therefore more easily oxidizable (5). Additionally, in vivo in mice, levels of F<sub>3</sub>-IsoPs in tissues such as heart were virtually undetectable at baseline but supplementation of animals with EPA markedly increased quantities up to 27.4 ± 5.6 ng/g heart. But, of particular note, we found that EPA supplementation also markedly reduced levels of arachidonate-derived F<sub>2</sub>-IsoPs mouse heart tissues by over 60% (*P* < 0.05). These observations are significant because F<sub>2</sub>-IsoPs are generally considered to be pro-inflammatory molecules associated with the pathophysiological sequelae of oxidant stress. It is thus intriguing to propose that part of the mechanism by which EPA prevents certain diseases is its ability to decrease F<sub>2</sub>-IsoP generation. In addition, it suggests that supplementation with fish oil may be of benefit to populations associated with increased levels of F<sub>2</sub>-IsoPs.

More recently, our laboratory has examined the formation of F-ring IsoPs generated from adrenic acid (C22:4, ω-6) (7). These compounds are termed F<sub>2</sub>-dihomo-IsoPs. Adrenic acid, like DHA, is highly enriched in the brain but is primarily found in white matter and is associated with myelin. White matter is commonly damaged by ischemic stroke and is uniformly damaged in multiple sclerosis. We reported that F<sub>2</sub>-dihomo-IsoPs are formed in significant amounts from adrenic acid and levels are markedly increased in settings of oxidant stress occurring in the white matter portion of the brain in humans. Proportionally, levels of dihomom-IsoPs in white matter undergoing oxidative injury increase to a greater extent than IsoPs and NPs derived from arachidonic acid and DHA, respectively. These studies suggest that quantification of dihomom-IsoPs may be a selective marker of white matter injury in vivo.

#### SUMMARY AND THOUGHTS FOR THE FUTURE

The discovery of the IsoPs as products of nonenzymatic lipid peroxidation has been a major contribution to lipid oxidation and free radical chemistry. Our understanding of the IsoP pathway continues to expand, providing new insights into the nature of lipid peroxidation in vivo and revealing new molecules that exert potent biological actions and might serve as unique indices of disease. Basic research into the biochemistry and pharmacology of the IsoPs, coupled with clinical studies employing these molecules as biomarkers, should continue to provide important insights into the role of oxidant stress in human physiology and pathophysiology. ■

#### REFERENCES

- Halliwell, B., and J. M. Gutteridge. 1990. Role of free radicals and catalytic metal ions in human disease: an overview. *Methods Enzymol.* **186**: 1–85.
- Morrow, J. D., K. E. Hill, R. F. Burk, T. M. Nammour, K. F. Badr, and L. J. Roberts 2nd. 1990. A series of prostaglandin F<sub>2</sub>-like compounds are produced in vivo in humans by a non-cyclooxygenase, free radical-catalyzed mechanism. *Proc. Natl. Acad. Sci. USA.* **87**: 9383–9387.
- Kadiiska, M. B., B. C. Gladen, D. D. Baird, D. Germolec, L. B. Graham, C. E. Parker, A. Nyska, J. T. Wachsman, B. N. Ames, S. Basu, et al. 2005. Biomarkers of oxidative stress study II: are oxidation products of lipids, proteins, and DNA markers of CCl<sub>4</sub> poisoning? *Free Radic. Biol. Med.* **38**: 698–710.
- Fam, S. S., and J. D. Morrow. 2003. The isoprostanes: unique products of arachidonic acid oxidation—a review. *Curr. Med. Chem.* **10**: 1723–1740.
- Gao, L., H. Yin, G. L. Milne, N. A. Porter, and J. D. Morrow. 2006. Formation of F-ring isoprostane-like compounds (F<sub>3</sub>-isoprostanes) in vivo from eicosapentaenoic acid. *J. Biol. Chem.* **281**: 14092–14099.
- Roberts 2nd, L. J., T. J. Montine, W. R. Markesbery, A. R. Tapper, P. Hardy, S. Chemtob, W. D. Dettbarn, and J. D. Morrow. 1998. Formation of isoprostane-like compounds (neuroprostanes) in vivo from docosahexaenoic acid. *J. Biol. Chem.* **273**: 13605–13612.
- VanRollins, M., R. L. Woltjer, H. Yin, J. D. Morrow, and T. J. Montine. 2008. F<sub>2</sub>-dihomo-isoprostanes arise from free radical attack on adrenic acid. *J. Lipid Res.* **49**: 995–1005.
- Taber, D. F., J. D. Morrow, and L. J. Roberts 2nd. 1997. A nomenclature system for the isoprostanes. *Prostaglandins.* **53**: 63–67.
- Rokach, J., S. P. Khanapure, S. W. Hwang, M. Adiyaman, J. A. Lawson, and G. A. FitzGerald. 1997. Nomenclature of isoprostanes: a proposal. *Prostaglandins.* **54**: 853–873.
- Yin, H., L. Gao, H. H. Tai, L. J. Murphey, N. A. Porter, and J. D.

- Morrow. 2007. Urinary prostaglandin F<sub>2</sub>alpha is generated from the isoprostane pathway and not the cyclooxygenase in humans. *J. Biol. Chem.* **282**: 329–336.
11. Stafforini, D. M., J. R. Sheller, T. S. Blackwell, A. Sapirstein, F. E. Yull, T. M. McIntyre, J. V. Bonventre, S. M. Prescott, and L. J. Roberts 2nd. 2006. Release of free F<sub>2</sub>-isoprostanes from esterified phospholipids is catalyzed by intracellular and plasma platelet-activating factor acetylhydrolases. *J. Biol. Chem.* **281**: 4616–4623.
  12. Morrow, J. D., J. A. Awad, H. J. Boss, I. A. Blair, and L. J. Roberts 2nd. 1992. Non-cyclooxygenase-derived prostanoids (F<sub>2</sub>-isoprostanes) are formed in situ on phospholipids. *Proc. Natl. Acad. Sci. USA.* **89**: 10721–10725.
  13. Morrow, J. D., T. A. Minton, K. F. Badr, and L. J. Roberts 2nd. 1994. Evidence that the F<sub>2</sub>-isoprostane, 8-epi-prostaglandin F<sub>2</sub> alpha, is formed in vivo. *Biochim. Biophys. Acta.* **1210**: 244–248.
  14. Morrow, J. D. 2006. The isoprostanes - unique products of arachidonate peroxidation: their role as mediators of oxidant stress. *Curr. Pharm. Des.* **12**: 895–902.
  15. Hou, X., L. J. Roberts 2nd, F. Gobeil, Jr., D. Taber, K. Kanai, D. Abran, S. Brault, D. Checchin, F. Sennlaub, P. Lachapelle, et al. 2004. Isomer-specific contractile effects of a series of synthetic f<sub>2</sub>-isoprostanes on retinal and cerebral microvasculature. *Free Radic. Biol. Med.* **36**: 163–172.
  16. Patrignani, P. 2003. Aspirin insensitive eicosanoid biosynthesis in cardiovascular disease. *Thromb. Res.* **110**: 281–286.
  17. Habib, A., and K. F. Badr. 2004. Molecular pharmacology of isoprostanes in vascular smooth muscle. *Chem. Phys. Lipids.* **128**: 69–73.
  18. Benndorf, R. A., E. Schwedhelm, A. Gnann, R. Taheri, G. Kom, M. Didie, A. Steenpass, S. Ergun, and R. H. Boger. 2008. Isoprostanes inhibit vascular endothelial growth factor-induced endothelial cell migration, tube formation, and cardiac vessel sprouting in vitro, as well as angiogenesis in vivo via activation of the thromboxane A<sub>2</sub> receptor. A potential link between oxidative stress and impaired angiogenesis. *Circ. Res.* **103**: 1037–1046.
  19. Milne, G. L., E. S. Musiek, and J. D. Morrow. 2005. F<sub>2</sub>-isoprostanes as markers of oxidative stress in vivo: an overview. *Biomarkers.* **10** (Suppl 1): S10–S23.
  20. Milne, G. L., H. Yin, J. D. Brooks, S. Sanchez, L. Jackson Roberts 2nd, and J. D. Morrow. 2007. Quantification of F<sub>2</sub>-isoprostanes in biological fluids and tissues as a measure of oxidant stress. *Methods Enzymol.* **433**: 113–126.
  21. Milne, G. L., S. C. Sanchez, E. S. Musiek, and J. D. Morrow. 2007. Quantification of F<sub>2</sub>-isoprostanes as a biomarker of oxidative stress. *Nat. Protoc.* **2**: 221–226.
  22. Pratico, D., O. P. Barry, J. A. Lawson, M. Adiyaman, S. W. Hwang, S. P. Khanapure, L. Iuliano, J. Rokach, and G. A. FitzGerald. 1998. IPF<sub>2</sub>alpha-I: an index of lipid peroxidation in humans. *Proc. Natl. Acad. Sci. USA.* **95**: 3449–3454.
  23. Mas, E., F. Michel, A. Guy, V. Bultel, Y. Falquet, P. Chardon, J. C. Rossi, J. P. Cristol, and T. Durand. 2008. Quantification of urinary F(2)-isoprostanes with 4(RS)-F(4t)-neuroprostane as an internal standard using gas chromatography-mass spectrometry application to polytraumatized patients. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **872**: 133–140.
  24. Liang, Y., P. Wei, R. W. Duke, P. D. Reaven, S. M. Harman, R. G. Cutler, and C. B. Heward. 2003. Quantification of 8-iso-prostaglandin-F(2alpha) and 2,3-dinor-8-iso-prostaglandin-F(2alpha) in human urine using liquid chromatography-tandem mass spectrometry. *Free Radic. Biol. Med.* **34**: 409–418.
  25. Bohnstedt, K. C., B. Karlberg, L. O. Wahlund, M. E. Jonhagen, H. Basun, and S. Schmidt. 2003. Determination of isoprostanes in urine samples from Alzheimer patients using porous graphitic carbon liquid chromatography-tandem mass spectrometry. *J. Chromatogr. B Analyt. Technol. Biomed. Life Sci.* **796**: 11–19.
  26. Taylor, A. W., R. S. Bruno, and M. G. Traber. 2008. Women and smokers have elevated urinary F(2)-isoprostane metabolites: a novel extraction and LC-MS methodology. *Lipids.* **43**: 925–936.
  27. Basu, S. 1998. Radioimmunoassay of 8-iso-prostaglandin F<sub>2</sub>alpha: an index for oxidative injury via free radical catalysed lipid peroxidation. *Prostaglandins Leukot. Essent. Fatty Acids.* **58**: 319–325.
  28. Morrow, J. D., Y. Chen, C. J. Brame, J. Yang, S. C. Sanchez, J. Xu, W. E. Zackert, J. A. Awad, and L. J. Roberts. 1999. The isoprostanes: unique prostaglandin like products of free radical-catalyzed lipid peroxidation. *Drug Metab. Rev.* **31**: 117–139.
  29. Morrow, J. D. 2005. Quantification of isoprostanes as indices of oxidant stress and the risk of atherosclerosis in humans. *Arterioscler. Thromb. Vasc. Biol.* **25**: 279–286.
  30. Davi, G., F. Chiarelli, F. Santilli, M. Pomilio, S. Vigneri, A. Falco, S. Basili, G. Ciabattoni, and C. Patrono. 2003. Enhanced lipid peroxidation and platelet activation in the early phase of type 1 diabetes mellitus: role of interleukin-6 and disease duration. *Circulation.* **107**: 3199–3203.
  31. Keaney, J. F., Jr., M. G. Larson, R. S. Vasani, P. W. Wilson, I. Lipinska, D. Corey, J. M. Massaro, P. Sutherland, J. A. Vita, and E. J. Benjamin. 2003. Obesity and systemic oxidative stress: clinical correlates of oxidative stress in the Framingham Study. *Arterioscler. Thromb. Vasc. Biol.* **23**: 434–439.
  32. Morrow, J. D., B. Frei, A. W. Longmire, J. M. Gaziano, S. M. Lynch, Y. Shyr, W. E. Strauss, J. A. Oates, and L. J. Roberts 2nd. 1995. Increase in circulating products of lipid peroxidation (F<sub>2</sub>-isoprostanes) in smokers. Smoking as a cause of oxidative damage. *N. Engl. J. Med.* **332**: 1198–1203.
  33. Montine, K. S., J. F. Quinn, J. Zhang, J. P. Fessel, L. J. Roberts 2nd, J. D. Morrow, and T. J. Montine. 2004. Isoprostanes and related products of lipid peroxidation in neurodegenerative diseases. *Chem. Phys. Lipids.* **128**: 117–124.
  34. Davi, G., G. Ciabattoni, A. Consoli, A. Mezzetti, A. Falco, S. Santarone, E. Pennese, E. Vitacolonna, T. Bucciarelli, F. Costantini, et al. 1999. In vivo formation of 8-iso-prostaglandin f<sub>2</sub>alpha and platelet activation in diabetes mellitus: effects of improved metabolic control and vitamin E supplementation. *Circulation.* **99**: 224–229.
  35. Roberts 2nd, L. J., J. A. Oates, M. F. Linton, S. Fazio, B. P. Meador, M. D. Gross, Y. Shyr, and J. D. Morrow. 2007. The relationship between dose of vitamin E and suppression of oxidative stress in humans. *Free Radic. Biol. Med.* **43**: 1388–1393.
  36. Gao, L., W. E. Zackert, J. J. Hasford, M. E. Danekis, G. L. Milne, C. Remmert, J. Reese, H. Yin, H. H. Tai, S. K. Dey, et al. 2003. Formation of prostaglandins E<sub>2</sub> and D<sub>2</sub> via the isoprostane pathway: a mechanism for the generation of bioactive prostaglandins independent of cyclooxygenase. *J. Biol. Chem.* **278**: 28479–28489.
  37. Montine, T. J., K. S. Montine, E. E. Reich, E. S. Terry, N. A. Porter, and J. D. Morrow. 2003. Antioxidants significantly affect the formation of different classes of isoprostanes and neuroprostanes in rat cerebral synaptosomes. *Biochem. Pharmacol.* **65**: 611–617.
  38. Chen, Y., J. D. Morrow, and L. J. Roberts 2nd. 1999. Formation of reactive cyclopentenone compounds in vivo as products of the isoprostane pathway. *J. Biol. Chem.* **274**: 10863–10868.
  39. Milne, G. L., G. Zanoni, A. Porta, S. Sasi, G. Vidari, E. S. Musiek, M. L. Freeman, and J. D. Morrow. 2004. The cyclopentenone product of lipid peroxidation, 15-A<sub>2t</sub>-isoprostane, is efficiently metabolized by HepG2 cells via conjugation with glutathione. *Chem. Res. Toxicol.* **17**: 17–25.
  40. Milne, G. L., L. Gao, A. Porta, G. Zanoni, G. Vidari, and J. D. Morrow. 2005. Identification of the major urinary metabolite of the highly reactive cyclopentenone isoprostane 15-A(2t)-isoprostane in vivo. *J. Biol. Chem.* **280**: 25178–25184.
  41. Brame, C. J., R. G. Salomon, J. D. Morrow, and L. J. Roberts 2nd. 1999. Identification of extremely reactive gamma-ketoaldehydes (isolevuglandins) as products of the isoprostane pathway and characterization of their lysyl protein adducts. *J. Biol. Chem.* **274**: 13139–13146.
  42. Yin, H., and N. A. Porter. 2005. New insights regarding the autoxidation of polyunsaturated fatty acids. *Antioxid. Redox Signal.* **7**: 170–184.
  43. Fessel, J. P., N. A. Porter, K. P. Moore, J. R. Sheller, and L. J. Roberts 2nd. 2002. Discovery of lipid peroxidation products formed in vivo with a substituted tetrahydrofuran ring (isofurans) that are favored by increased oxygen tension. *Proc. Natl. Acad. Sci. USA.* **99**: 16713–16718.
  44. Roberts 2nd, L. J., and J. P. Fessel. 2004. The biochemistry of the isoprostane, neuroprostane, and isofuran pathways of lipid peroxidation. *Chem. Phys. Lipids.* **128**: 173–186.
  45. Patel, M., L. P. Liang, H. Hou, B. B. Williams, M. Kmiec, H. M. Swartz, J. P. Fessel, and L. J. Roberts 2nd. 2008. Seizure-induced formation of isofurans: novel products of lipid peroxidation whose formation is positively modulated by oxygen tension. *J. Neurochem.* **104**: 264–270.
  46. Serhan, C. N., C. B. Clish, J. Brannon, S. P. Colgan, N. Chiang, and K. Gronert. 2000. Novel functional sets of lipid-derived mediators with antiinflammatory actions generated from omega-3 fatty acids via cyclooxygenase 2-nonsteroidal antiinflammatory drugs and transcellular processing. *J. Exp. Med.* **192**: 1197–1204.
  47. Pratico, D., E. M. Smyth, F. Violi, and G. A. FitzGerald. 1996. Local amplification of platelet function by 8-Epi prostaglandin F<sub>2</sub>alpha is not mediated by thromboxane receptor isoforms. *J. Biol. Chem.* **271**: 14916–14924.